

Effect of Increased Creatine Kinase Levels in Rabbits Due to Phospholipase A₂ of *Naja nigricollis* Venom

*¹Kurfi BG, ²Abdulazeez MA, ³Tukur Z

¹Department of Biochemistry, Faculty of Biomedical Sciences, Bayero University, Kano, Nigeria

²Department of Biotechnology, Bayero University, Kano, Nigeria

³Department of Biological Sciences, Faculty of Science, Bayero University, Kano, Nigeria

*Corresponding Author: Kurfi BG, Department of Biochemistry,
Faculty of Biomedical Sciences, Bayero University, Kano, Nigeria
Email: binkurfi@gmail.com*

Abstract

Snakebite envenomation is an important public health problem among rural communities in northern Nigeria. Phospholipase A₂ was isolated from the venom of *Naja nigricollis* by a combination of gel filtration on Sephadex G-75 and ion-exchange chromatography on DEAE cellulose. Sodium dodecyl sulfate-polyacrylamide gel filtration showed the enzyme to be a dimer with molecular weight of 65 kDa. The effect of the enzyme Phospholipase A₂ when administered intramuscularly (1.00-5.00µg/g at a given volume of 50µl) exhibited increase in plasma Creatinine Kinases activity that resulted in Myonecrosis due to intense destruction of muscular fiber which involves local infiltration of inflammatory cells that was dose and time-dependent. Rabbits (n=16) were grouped into four according to the dosage and time. Serum CK activity increased from a lower level at different time of 1,3 and 6 hours after envenomation 120.22 ± 10.83 IU/L to 2281.80 ± 609.43 IU/L. Necrosis was more extensive when higher doses of Phospholipase A₂ were injected and also, there was a corresponding increase in serum CK levels from 382.31 ± 122.70 to 818.31 ± 133.32 IU/L progressively. These findings suggest that Phospholipase A₂ induce acute muscle cell damage and oedema upon intramuscular injection in rabbit, as evidenced by increase in serum CK activity. Also, CK was measured as an index of myotoxicity. This findings provide a useful molecular basis in improved treatment and management of envenomation by *Naja nigricollis*.

Keywords: *Naja nigricollis*, Phospholipase A₂, Creatine Kinase.

Introduction

The venom of Black-Neck Spitting Cobra snake, *Naja nigricollis* is known to induce many toxic effects that may lead to death of the victim. In Nigeria, more severe cases of snakebite envenomation are inflicted by cobra species of the family elapidae. Surveys conducted in the more densely populated northern savanna show that *Naja nigricollis* is the predominant species of medical importance. It is a major medical problem among rural communities of the savanna region of West Africa, notably in Benin, Burkina-Faso, Cameroon, Ghana, Nigeria and Togo. West Africa bears the heaviest burden of snakebite, with an estimated incidence of 500 bites and 50 deaths/100,000 population reported in rural Nigeria alone.^[1]

Snake venoms are particularly rich in s and contain multiple forms of Phospholipase A₂ (PLA₂) enzymes,^[2]

that are similar in molecular size, isoelectric points and even amino terminal sequences.^[3] However, (PLA₂) isoenzymes may exhibit a specific pharmacological effect, such as pre-synaptic or post-synaptic neurotoxicity, cardiotoxicity, myotoxicity, platelet aggregation, oedema formation, haemolysis, anticoagulation, convulsion, and hypotension.^[4,5]

PLA₂ from snake venoms exhibit a wide variety of pharmacological effects by interfering with normal physiological processes.^[6] PLA₂ triggers a cascade of inflammatory events characterized by increased microvascular permeability and oedema formation, leukocyte recruitment into tissues, nociception and release of inflammatory mediators which mimic a number of systemic and local inflammatory disorders in humans.^[7]

Information on the effect of Creatine Kinase and Phospholipase A₂ from *Naja nigricollis* venom found in this locality and its potential role in the pathophysiology of envenomation is scanty. It is well established that the analyses of serum enzymes can help to detect some abnormalities in the liver and heart.

This study is aimed to assess the effect of isolated enzyme at different time interval and, doses of Phospholipase A₂ on Creatine Kinase from African black-necked spitting cobra (*Naja nigricollis*) venom.

Materials and Methods

Chemicals

General laboratory and inorganic chemicals were obtained from Aldrich Chemical Company, and were of analytical grade.

Animals

Sixteen young healthy rabbits (300-400kg), seven months old were purchased from the Department of Biological Sciences, Faculty of Science, Bayero University, Kano. The animals were de-wormed and allowed to acclimatize for a week before the commencement of the experiment.

Naja nigricollis was caught from the wild at Gombe in the northern part of Nigeria. It was identified in the Zoology unit of the Biological Sciences Department of Ahmadu Bello University, Zaria, Nigeria.

Purification of PLA₂ Enzyme

The *N. nigricollis* (200 mg) venom dissolved in 1 ml equilibration buffer (0.05 M Tris-HCl, pH 6.8), was loaded on Sephadex G-75 column (2.6 × 50 cm). The sample-dissolving buffer was used for equilibration of the Sephadex column, and elution of the loaded samples. Fractions of 10 ml were collected at a flow rate of 48 ml/hr using fraction collector (Pharmacia LKB, Sweden). Fractions with phospholipase A₂ activity recovered from the previous step were pooled and directly applied to a DEAE Cellulose column (1.6 × 25 cm) pre-equilibrated with the same buffer and eluted with a linear NaCl gradient from 0 to 1.3 M in the same buffer.

Phospholipase A₂ Activity

Phospholipase A₂ activity was performed as described by Dole (30). Briefly, crude egg yolk phospholipids

(diluted 1:5 in 0.1 M Tris-HCl, 0.01 M CaCl₂, pH 8.5) were used as substrate in the presence of 1% Triton X-100. The isolated venom (5, 25 and 50 µg) was added to 10 mL of substrate, incubated at 37°C for 15 minutes, and then the free fatty acids were extracted and titrated with 0.018 M NaOH. Crude venom (10 µg) was utilised as a positive control.

Polyacrylamide Gel Electrophoresis (SDS-PAGE)

The electrophoretic analysis was performed in the Mini-Protean II Dual- Slab Cell (Bio Rad). Preparation of gels, samples, and electrophoresis were performed according to the conditions described by Laemmli^[9]. Proteins were stained with 0.2% Coomassie Brilliant Blue R-250 in fixative solution.

In-vivo Myotoxicity

Groups 2-8 rabbits were injected i.m. with different dose (1.00-5.00 µl/kg) of isolated enzyme Phospholipase A₂ dissolved in 0.1 ml of phosphate buffered saline solution, pH 7.2. Three mice were used as control samples, receiving each of them 0.1ml/kg of phosphate buffered saline solution (pH 7.2). After 1, 3, 6 hrs of PLA₂ enzyme injection, rabbits were anaesthetised with chloroform and blood samples were taken from the tail without using anticoagulant. Serum was obtained for the enzymatic determination of Creatine Kinase (CK) levels using a kinetic method U. V. (Randox). The CK activity was expressed in international units per liter, with one unit defined as the amount of enzyme that transfers 1.0 mM of phosphate from phosphocreatine to ADP per min at pH 7.4 at 30°C. Creatine Kinase activity was expressed in units per litre

Blood Sample Collection

Sixteen rabbits were anaesthetized and blood samples were collected. Stored venoms were dissolved in 10 ml of normal saline solution and were injected intramuscularly to the rabbits (300 µg/ kg body weight of rabbit) and their sera at different dose were collected. Blood samples were centrifuged (Sigma centrifuge, Model 6k15) for 10 min at 2500 rpm. Quantitative detection of CK are determined with the help of commercially available kits (Sigma). The results are mean ±SD of three experiments.

Result

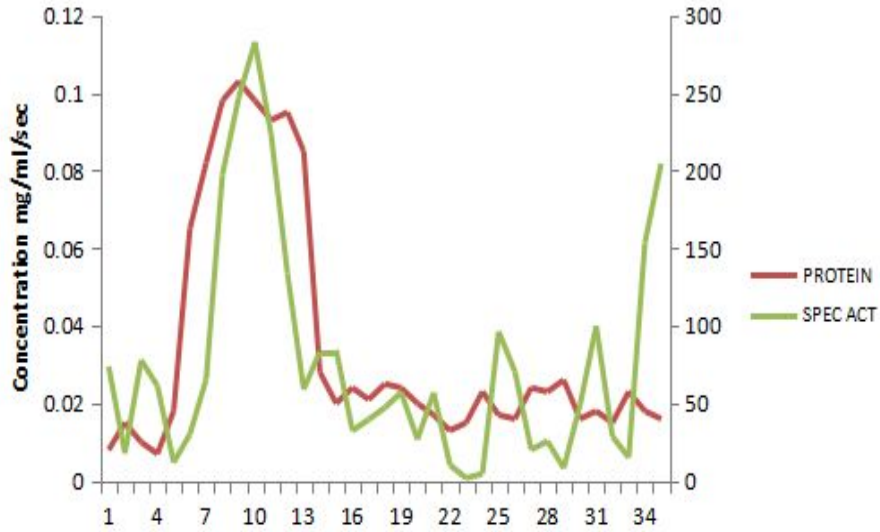


Figure I. Elution profile of *Najanigracollis* Phospholipase A₂ Using Sephadex G75

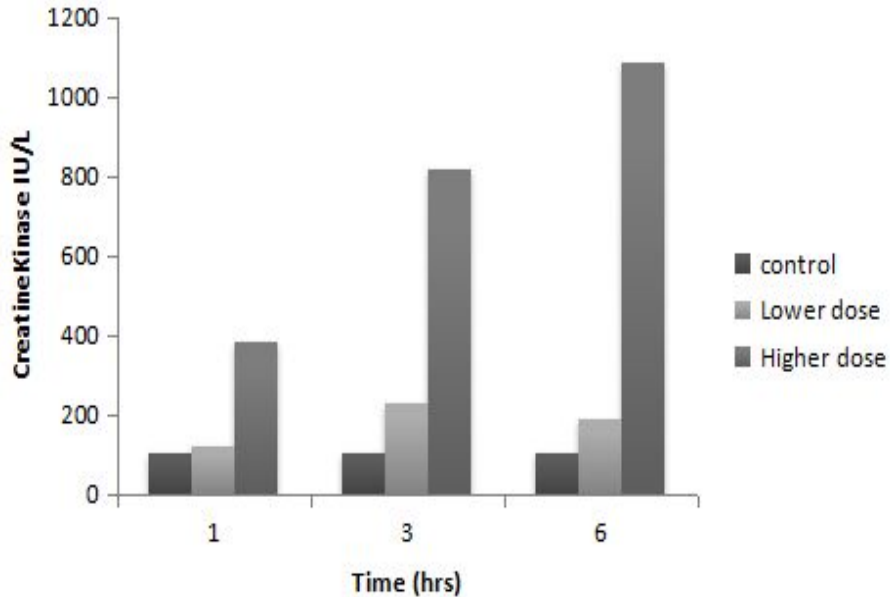


Figure2. Effect of dose and time on in vivo myotoxicity of *Naja nigricollis* venom in Rabbit. The venom phospholipase A₂ dose was increased from 1.0 to 5.0µg/g, From 1.00- 2.5µg/g was referred as the lower dose and 3.00-5.00 µg/g as the higher dose. Plasma Creatinine Kinase was measured after 1, 3.and 6hrs after the venom injection.

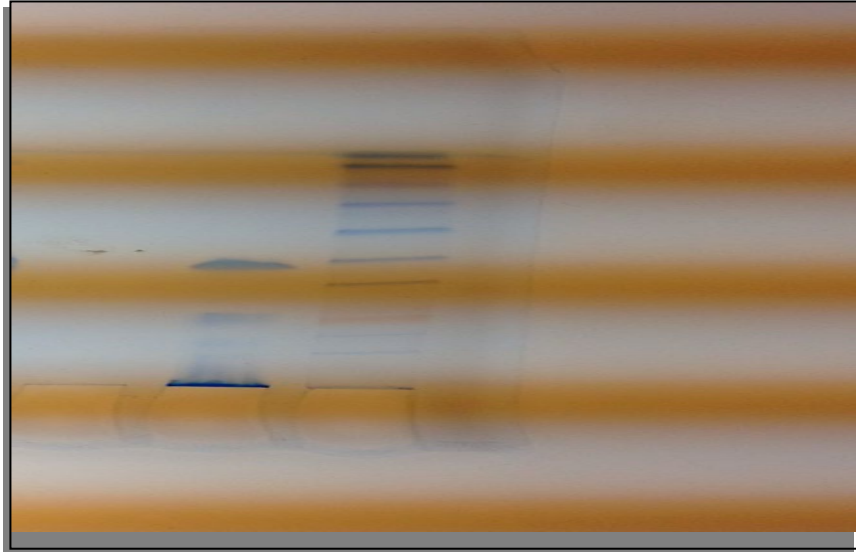
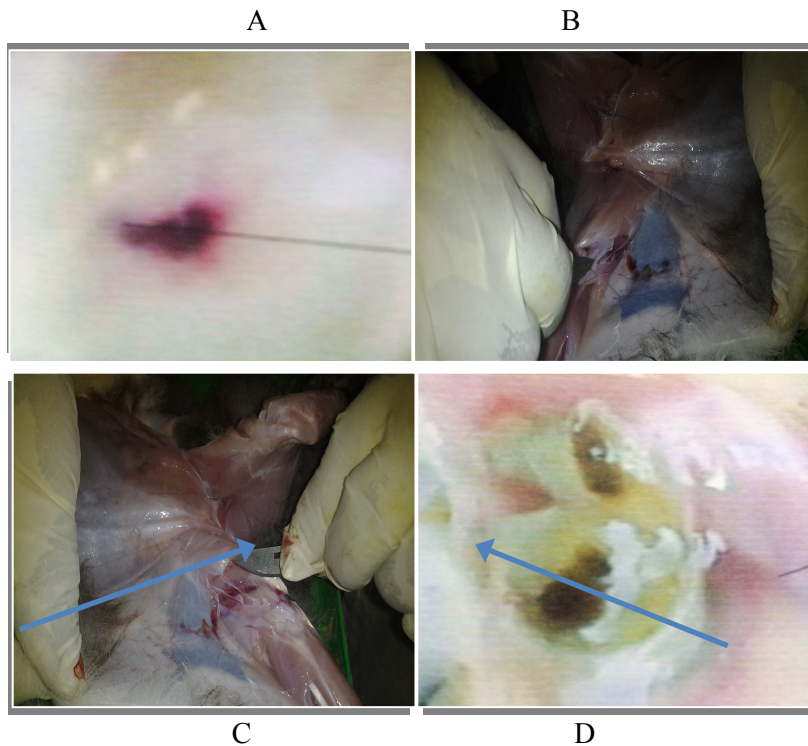


Plate1: Protein Electrophoregrams and Phospholipase A₂ Activity of Purified *N. nigricollis* Key: Lane 1=molecular weight marker, Lane 2=purified enzyme (65KDa),



Dnecrosis

Plate 2 ABCD: A slight haemorrhage when injected (10µl)with the isolated venom (phospholipase A₂) Plate B and C indicated damage of the muscular tissue with a dose of (20µl) after 3hrs. Necrosis and haemorrhagic severe tissue damage at plate D with the dose of (50µl) after 6hrs of intramuscular injection.

Discussion

Phospholipase A₂ was isolated from *N. Nigricollis* snake venom. It was purified to homogeneity by only two simple chromatographic steps. This phospholipase exhibited high edematogenic and myotoxic activities, demonstrating its capacity to contribute to tissue damage after snakebite or envenoming produced by this *Naja nigricollis* venom. The two highest peaks from (figure 1) indicated the increase in specific activity of the crude PLA² after the two purification steps could be due to the removal of other synergistically interacting components of the venom,^[10] these two peaks were selected for further characterization of Phospholipase A₂.

The enzyme appeared as a single band at 65 kDa in SDS PAGE, after reduction by 2-mercaptoethanol (plate 1) which agreed with Najanaja.^[11] Indeed, myotoxins of snake venoms affect mainly the plasma membrane of muscle cells to which they bind through their cationic sequence.^[12] Molecular mechanism by which they caused the muscle tissue damage is not yet fully elucidated. Myonecrosis is due to the myotoxins that induce irreversible damage of skeletal muscle fibers. These molecules bind to the plasma membrane of muscle cells and alter its permeability and integrity (Plate 5). The toxin induced a dose and time dependent myonecrosis upon IM injection in Rabbit, as evidenced by the

significant increase in plasma CK activity (plate 2B and C).

Creatinine Kinase level increased since the first hour after injection to the sixth hour with the highest level (Plate A,B,C and D), the morphological characteristic of myonecrosis induced by this toxin are in distinguishable from those previously described for B.aspermyotoxin II,^[13 16] suggesting a common mechanism of action. Figure 2 shows the effect of increasing dose of the *Naja nigricollis* venom on plasma CK level, 382.31 ± 122.70 to 818.31 ± 133.32 IU/L, these results indicate that the plasma CK levels depend on the dose and time of venom induction. Several studies show that PLA² enzyme of *bothropic* venom are also able to increase the plasma CK level.^[14, 15] These data suggest that increasing concentration of venom result in increasing muscles lesion, and further demonstrate that increasing dose of the injection can also increase these plasma CK level.

Conclusion

The phospholipase A₂ exhibited high edematogenic and myotoxic activities, demonstrating its capacity to contribute to tissue damage after snakebite and envenoming produced by the *Naja nigricollis* snake.

Acknowledgement

The authors would like to thank Venom Antivenom research Group of Bayero University Kano for the financial support to conduct the research.

Reference

- Habib AG, Gebi UI, Onyemelukwe GC. Snake bite in Nigeria. *African Journal of Medicine and medical Sciences* 2001; **30**, 171-178. [Review]
- Daniele JJ, Bianco ID, Fidelio GD (1995). Kinetic and pharmacologic characterisation of phospholipases A₂ from *Bothrops neuwiedii* venom. *Arch BiochemBiophys.* **318**:65-70
- Kini RM. Excitement ahead: structure, function and mechanism of snake venom phospholipase A₂ enzymes. *Toxicon*2003;**42**:827-40.
- Santos-Filho NA, Silveira LB, Oliveira CZ, Bernardes CP, Menaldo DL, Fuly AL. A new acidic myotoxic, anti-platelet and prostaglandin I₂ inductor phospholipase A₂ isolated from *Bothrops moojeni* snake venom. *Toxicon.* 2008;**52**(8):908-17.
- Zouari-Kessentini R, Srairi-Abid N, Bazaa A, El Ayeb M, Luis J, Marrakchi N.; Antitumoral potential of Tunisian Snake Venoms secreted phospholipases A₂. *J.Comp.Physio* 2013;**2**(1) 544-546.
- Lambeau G, Gelb MH. Biochemistry and physiology of mammalian secreted phospholipases A₂. *Annu. Rev. Biochem.*2008; **77**, 495-520.

- 7 Kamiguti AS, Cardoso JI, Theakson RD, Sano-Martius IS, Hutton RA, Rugman FP, Warrell DA, Hay CR. Coagulopathy and haemorrhage in human victims of *Bothrops jararaca* envenoming in Brazil. *Toxicon* 1991;**29**:961-72.
- 8 Bradford MM. A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein-dye binding. *Analys. Biochem.* 1979;(72):248-54.
- 9 Laemmli UK. Cleavage of structural protein during the assembly of the head of bacteriophage T4. *Nature*. 1970;**227**(5259): 680-5.
- 10 Santos-Filho NA, Silveira LB, Oliveira CZ, Bernardes CP, Menaldo DL, Fuly AL. A new acidic myotoxic, anti-platelet and prostaglandin I₂ inductor phospholipase A₂ isolated from *Bothrops moojeni* snake venom. *Toxicon*. 2008;**52**(8):908-17.
- 11 Dole VP. Relation between non-esterified fatty acids in plasma and the metabolism of glucose. *J Clin Invest.* 1956;**35**(2):150-4.
- 13 Nakada K, Nakada F, Ito E, Inoue F. 1984. Quantification of myonecrosis and comparison of necrotic activity of snake venoms by determination of creatine phosphokinase activity in mice sera. *Toxicon*, **22**: 921-930.
- 14 Selistre HS, White SP, Ownby CL. cDNA cloning and sequence analysis of a lysine-49 phospholipase A₂ myotoxin from *Agkistrodon contortrix atrox* snake venom. *Arch. Biochem. Biophys.* 1996; 326:21
- 15 Ketelhut DF, de Mello MH, Veronese EL, Esmeraldino LE, Murakami MT, Arni RK, Giglio JL, Cintra AC, Sampalo SV. Isolation, characterisation and biological activity of acidic phospholipase A₂ isoforms from *Bothrops jararacussu* snake venom. *Biochimie* 2003;**85**:983-91.